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# Vitamin D metabolites across the menstrual cycle: a systematic review

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## Abstract

**Background:** Accurate estimation of vitamin D status is important for health research and can impact prevention and treatment of deficiency in women of reproductive age. We aimed to assess if blood concentrations of 25-hydroxyvitamin D [25(OH)D] or 1,25-dihydroxyvitamin D [1,25(OH)<sub>2</sub>D] change across the menstrual cycle.

**Methods:** We conducted a systematic search in PubMed, Web of Science, CAB and BIOSIS of literature published until December 2018 which reported concentrations of vitamin D metabolites at two or more identified points among women with regular menstrual cycles.

**Results:** Ten longitudinal studies met the inclusion criteria; nine studies measured 1,25(OH)<sub>2</sub>D and five studies measured 25(OH)D. Study size ranged from 5 to 47 subjects, with an age range of 18–47 years. One study found a decrease in concentration of 25(OH)D in the periovulatory and luteal phase. Four studies found no changes in concentrations of 25(OH)D. Two studies found a rise in 1,25(OH)<sub>2</sub>D within the follicular phase, including a 128% increase from day 1 to 15 and a 56% increase from day 0 to 12. Two studies found rises in 1,25(OH)<sub>2</sub>D concentrations from the follicular to luteal phase of 13 and 26%. Five studies did not find any changes in concentrations of 1,25(OH)<sub>2</sub>D.

**Conclusions:** No conclusion can be drawn on the pattern of 1,25(OH)<sub>2</sub>D concentrations across the normal menstrual cycle due to inconsistencies in study findings. Evidence is currently insufficient to assess 25(OH)D concentrations across the cycle. Future studies should aim to measure 1,25(OH)<sub>2</sub>D and 25(OH)D longitudinally, to understand relationships with other hormones and the potential impact on estimates of vitamin D deficiency.

**Keywords:** Menstrual period, Menstruation, Calcidiol, Calcitriol, Hydroxycholecalciferol, Reproductive hormones

## Background

Vitamin D deficiency has become a problem of public health concern, with increased prevalence among women of reproductive age and higher rates of deficiency in women compared to men [1–4].

Estimating vitamin D status accurately is important for public health surveillance and research, and can impact the ability to monitor and improve health outcomes across populations. The Institute of Medicine (IOM) defines vitamin D deficiency as having concentrations of 25-hydroxyvitamin D [25(OH)D] < 30 nmol/l (12 ng/ml) and vitamin D insufficiency as 25(OH)D < 50 nmol/l (20 ng/ml) [5]. Over the last decade, the prevalence of vitamin D insufficiency was high among

adult women living in Bangladesh (80%), Sri Lanka (59%), Israel (51–60%), and Germany (58%) [2]. Low vitamin D status is also a concern among women of reproductive age living in the US [4, 6, 7] with a quarter of women being classified as vitamin D insufficient [7]. Of concern, the prevalence of vitamin D deficiency was observed to be higher among non-Hispanic black women (41%) compared to non-Hispanic white women (4%) in the US [7].

Vitamin D from both cutaneous production and oral intake gets metabolized by the liver into 25(OH)D, the major circulating metabolite and primary indicator used for measurement of vitamin D status [8–10]. 25(OH)D is further metabolized in the kidneys to 1,25 dihydroxyvitamin D [1,25(OH)<sub>2</sub>D], the biologically active form of vitamin D that acts as a ligand for its nuclear receptor to promote genetic expression [8–10].

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Sources of vitamin D include synthesis in skin during sunlight exposure (specifically to ultraviolet B (UVB) radiation), food, and dietary supplements [11–14]. Salmon, mackerel, cod liver oil, shiitake mushrooms and egg yolk are among the few foods naturally containing vitamin D, leaving skin production, vitamin D fortified foods, or supplements as the major source for most of the world [8, 11, 14]. Many factors increase the risk of vitamin D deficiency including poor dietary intake; vegan diet; low availability of fortified foods; religious and cultural practices that reduce sun exposure; darker skin pigmentation; and living in higher latitudes without UVB radiation in the winter [13, 15, 16].

Vitamin D deficiency has been associated with increased risk of diabetes, cardiovascular disease, cognitive decline, certain types of cancer, depression [9, 11], and adverse pregnancy outcomes [17–22]. Poor vitamin D status has also been associated with early menarche, dysmenorrhea, premenstrual syndrome (PMS), and uterine fibroids [23], mechanistically plausible through vitamin D receptors expressed in the ovary, placenta, and uterus [23–26].

The female menstrual cycle is associated with a series of changes occurring in the uterus and ovaries of females resulting in ovulation and the shedding of the uterine lining when conception does not occur [27, 28]. It is driven by the interaction of several hormones produced by the hypothalamus, anterior pituitary, and ovaries, including follicle stimulating hormone (FSH), luteinizing hormone (LH), estrogen, and progesterone [27, 28]. The menstrual cycle begins with menses and is broadly divided into the follicular (pre-ovulation) and luteal phases (post-ovulation) [27, 28].

Concentrations of micronutrients have been reported to change during the menstrual cycle [29, 30]. Zinc has been reported to vary across the menstrual cycle at different time points with the highest concentrations observed at ovulation [29]. Similarly, cyclic changes in copper have been reported with the highest concentrations occurring during menses; and zinc and copper changes might each be due to changing concentrations of estrogen [29]. Biomarkers used to assess iron status such as hemoglobin, transferrin saturation, and ferritin have also been reported to change during the menstrual cycle with the highest concentrations observed during the luteal phase and lowest during menstruation [30]. These changes are thought to result from variations in ovarian hormones and/or plasma volume [30].

While it is known that 1,25(OH)<sub>2</sub>D concentrations increase two- to three-fold during pregnancy [31], and there are some reports of 25(OH)D and 1,25(OH)<sub>2</sub>D across the menstrual cycle, to the authors' knowledge these biomarkers have not been well chronicled pre-pregnancy. Because concentrations of several nutrition biomarkers change across the menstrual cycle [29, 30,

32], and because vitamin D is a hormone, we hypothesized that vitamin D metabolites may also have cyclic changes along with reproductive hormones. Thus, our objective was to conduct a systematic review of studies that assessed concentrations of 25(OH)D and/or 1,25(OH)<sub>2</sub>D longitudinally to assess if these vitamin D biomarkers change across the normal menstrual cycle.

## Methods

### Eligibility criteria

Original studies which measured concentrations of 25(OH)D and/or 1,25(OH)<sub>2</sub>D in serum or plasma at two or more identified time points per woman were eligible for inclusion in the review. We included studies of normal, healthy women with regular menstrual cycles (26–35 days), and articles written in English. We excluded any review articles or studies which examined women with a specific health issue (e.g. PMS) or among those with irregular cycles or women who were taking any sort of medication. However, we did not exclude studies among women taking oral contraceptives because this is such a common practice (although notably it introduces difficulty in accurately estimating the phases of the menstrual cycle). If a study included a healthy group that met our criteria and any other group outside the criteria, we included data from the healthy group only. Inclusion of studies was not limited by publication date. All laboratory methods for measurement of vitamin D metabolite concentrations were accepted.

### Search strategy

We conducted a systematic search of literature in PubMed, Web of Science, CAB and BIOSIS of literature published until December 2018. The search strategy used several combinations of keywords and MeSH terms. The search strategy used for PubMed was (“menstrual cycle”[TIAB] OR “menstrual cycle”[Mesh]) AND (“vitamin D”[TIAB] OR “vitamin D”[Mesh] OR “25-hydroxyvitamin D”[TIAB] OR “micronutrients”[TIAB] OR “micronutrients”[Mesh] OR “Hydroxycholecalciferols”[TIAB] OR “Hydroxycholecalciferols”[Mesh] OR “calcitriol”[Mesh] OR “calcitriol”[TIAB]). References in identified papers were also scanned for studies that met eligibility criteria. The articles were initially screened on the basis of title and abstract. After excluding irrelevant articles, full text of the remaining studies was screened for eligibility.

### Data collection

Information and variables of interest were extracted from all eligible studies. The main outcomes of interest were concentrations of 25(OH)D and 1,25(OH)<sub>2</sub>D and changes in these vitamin D metabolites across the menstrual cycle. Values were converted to ng/ml for concentration of 25(OH)D and to pg/ml for 1,25(OH)<sub>2</sub>D if reported otherwise.

This systematic review was conducted according to PRISMA (Preferred Reporting Items for Systematic Reviews and Meta-Analysis) recommendations [33].

**Results**

**Study selection**

We initially identified 376 studies which were retrieved using the search strategies; after removing duplicates, 238 studies remained (Fig. 1). Among these, 220 studies were not relevant on the basis of the title and abstract and were excluded. The remaining 18 studies underwent full review. Eight of these studies did not meet the eligibility criteria and were excluded, leaving ten unique, longitudinal studies included in this systematic review. Five studies measured the concentrations of 1,25(OH)<sub>2</sub>D alone [34–38], one study measured the concentration of 25(OH)D alone [39], and four studies measured the concentrations of both vitamin D metabolites [40–43].

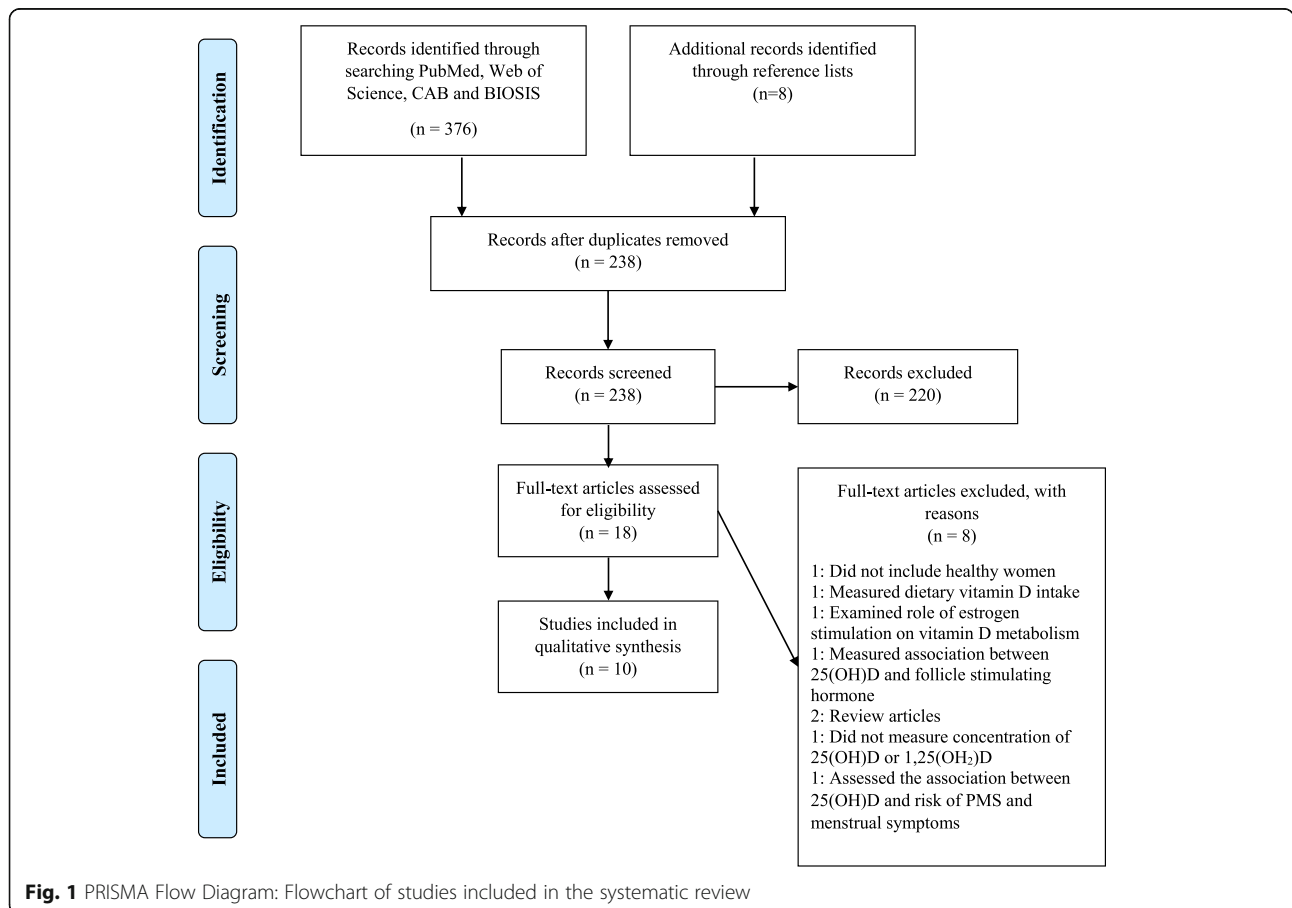
**Study characteristics**

Six studies were conducted in the US [34–37, 40, 41], two in Denmark [38, 42], one in Germany [43], and one in Ireland [39]. The study size ranged from 5 to 47 women and the age range across all studies was 18–47

years. The subjects included in the studies were typically a convenience sample of volunteers. Phases of the menstrual cycle were identified by the last menstrual period (LMP) date and/or hormonal evaluation. Eight studies measured vitamin D metabolites among healthy young women with normal menstrual cycles [34–39, 42, 43] and the remaining two studies measured metabolites among both healthy controls and women with PMS [40, 41].

The primary objective of the studies was the following: seven studies aimed to assess the concentrations of vitamin D metabolites and other calciotropic hormones and markers of bone metabolism across the menstrual cycle; one study aimed to assess the association between serum parathyroid hormone (PTH) and changes in endogenous estrogen; one study aimed to assess the effect of estrogens on calcium regulating hormones; and one study aimed to examine the metabolic changes across the menstrual cycle by performing hormonal and nutrient evaluation. All ten studies took at least one measurement within the follicular phase [34–43] and eight studies took measurements that spanned both follicular and luteal phases [35–41, 43].

The summary of findings for the studies are shown in Table 1. Tjellesen et al. [42] reported that the concentrations of 25(OH)D did not change within the follicular



**Table 1** Characteristics of studies which measured vitamin D metabolites longitudinally across the normal menstrual cycle in healthy women<sup>1</sup>

Author (Year)	N	Age range (years)	Method (Sample Type)	Mean (SD)25(OH)D (ng/ml)	Mean (SD)1,25(OH) <sub>2</sub> D (pg/ml)	Main Findings
Baran (1980) [34]	12	19–34	RLRA (Serum)	NA	EF (Day 3): 47 (3) <sup>4</sup> LF (Day 13): 51 (3)	No change in concentrations of 1,25 (OH) <sub>2</sub> D from day 3 to day 13
Gray (1982) [35]	7	18–35	RIA (Serum)	NA	EF (Day 1): 53.4 (11.8) <sup>a,4</sup> EF (Day 8): 56.3 (12.6) <sup>a</sup> EL (Day 15): 121 (20.1) <sup>b</sup> LL (Day 22): 86.9 (27.2)	Increase in concentration of 1,25(OH) <sub>2</sub> D from day 1 to day 15 Concentration of 1,25(OH) <sub>2</sub> D at midcycle was higher than days 1 and 8
Tjellesen (1983) [42]	5	23–29	PBA <sup>2</sup> UV-detection <sup>3</sup> (Serum)	NR	EF <sup>5</sup> : Mean change: 56% from day 2 to day 14	No change in concentration of 25(OH)D from day 2 to day 14 Concentration of 1,25(OH) <sub>2</sub> D increased from day 2 to day 14
Buchanan (1986) [36]	20	24–44	RRA (Serum)	NA	EF: 34 (3) <sup>a</sup> LF: 39 (3) EL: 43 (3) <sup>b</sup> LL: 37 (2)	A rise in the concentration of 1,25(OH) <sub>2</sub> D from the EF to EL phase
Muse (1986) [37]	6	26–39	CRA (Serum)	NA	All: 40.1 (1.7) <sup>4,6</sup>	A slight, non-statistically significant change in 1,25(OH) <sub>2</sub> D across the cycle
Nielsen (1990) [38]	8	20–47	RIA (Serum)	NA	EF: 56.5 (13.5) LF: 57.3 (21.9) EL: 57.6 (14.2) LL: 56.9 (10.0)	No change in concentrations of 1,25 (OH) <sub>2</sub> D across the cycle
Thys-Jacobs (1995) [40]	5	28–45	RRA <sup>2</sup> RBA <sup>3</sup> (Serum)	EF (Day 2) Midcycle (Days 12,13,14,15) LL (Day 26) All: 45.6 (12.8) <sup>7</sup> EF: NR Midcycle: 44.8 (11.6) LF: NR	All: 38.0 (8.8) <sup>7</sup> EF (Day 2): 30.3 (8.4) Midcycle (Days 12,13,14,15): 38.8 (14.6) LL (Day 26): 37.3 (8.8)	No change in concentrations of 25(OH)D across the cycle Statistical significance not reported for concentrations of 1,25(OH) <sub>2</sub> D across the cycle
Zittermann (2000) [43]	10	25 <sup>8</sup>	RRA <sup>2</sup> HPLC <sup>3</sup> (Serum)	EF: 10.3 (3.1) <sup>c</sup> LF: 9.3 (2.6) EL: 9.3 (2.4) ML: 8.8 (2.6) EF (next cycle): 8.4 (3.4) <sup>d</sup>	EF: 17.5 (4.1) LF: 19.4 (5.5) EL: 19.6 (5.1) ML: 18.4 (5.3) EF (next cycle): 17.1 (4.9)	No changes in 25(OH)D within the same cycle; Significant decrease in 25(OH)D from EF of one cycle to EF phase of next cycle No change in concentrations of 1,25(OH) <sub>2</sub> D across the cycle
Thys-Jacobs (2007) [41]	47	18–45	RRA <sup>2</sup> CLA <sup>3</sup> (Serum)	EF (Days 2 and 7) <sup>9</sup> Midcycle (Days 12,13,14,15) ML to LL (Days 22 and 27) <sup>10</sup> All: 26.1 (10.8) <sup>11</sup>	EF (Days 2 and 7) Midcycle (Days 12,13,14,15) ML to LL (Days 22 and 27) All: 49.0 (10.7) <sup>11</sup> EF: 46.4 <sup>a,12</sup> LL: 52.1 <sup>b,12</sup>	No change in concentrations of 25(OH)D across the cycle Significant rise in concentrations of 1,25(OH) <sub>2</sub> D from EF to LL phase
Draper (2018) [39]	34	19–39	CMIA	MP, FP, PO, LP and PP: NR	NA	Significant decrease in concentration of 25(OH)D in the periovulatory and luteal phase

CRA cytoreceptor assay, CLA chemiluminescence assay, CMIA chemiluminescent microparticle immunoassay, EF early follicular, EL early luteal, FP follicular phase, HPLC high-performance liquid chromatography, LF late follicular, LL late luteal, LP luteal phase, ML mid luteal, MP: menstrual phase; NA not applicable, NR not reported, PBA protein binding assay, PO periovulatory, PP pre-menstrual phase, RBA radiobinding assay, RIA radioimmunoassay, RLRA radioligand receptor assay, RRA radioreceptor assay

<sup>1</sup>All studies were conducted in the US except Tjellesen (1983) and Nielsen (1990) conducted in Denmark; Zittermann (2000) conducted in Germany and Draper (2018) conducted in Ireland

<sup>2</sup>Method used to measure concentration of 1,25(OH)<sub>2</sub>D

<sup>3</sup>Method used to measure the concentration of 25(OH)D

<sup>4</sup>Values are presented as mean (SE); SE, Standard Error

<sup>5</sup>Early follicular phase and days with highest concentration of estradiol (number of measurements not reported)

<sup>6</sup>Mean for all measurements across the cycle; measured daily throughout the cycle

<sup>7</sup>Mean for all measurements across the cycle

<sup>8</sup>Mean age reported

<sup>9</sup>Measurement for early follicular phase is approximately between day 6 to 2 days before ovulation

<sup>10</sup>Measurement for late luteal phase is approximately day 27 but varies according to the individual's cycle

<sup>11</sup>Mean of measurement obtained after determining luteinizing hormone (LH) surge

<sup>12</sup>Standard deviation not reported

<sup>a,b</sup>Concentrations of 1,25(OH)<sub>2</sub>D are significantly different between two time points

<sup>c,d</sup>Concentrations of 25(OH)D are significantly different between two time points

phase. Both studies by Thys-Jacobs and colleagues [40, 41] reported no changes in concentrations of 25(OH)D from the early follicular to the late luteal phase. Zittermann et al. [43] reported no changes in 25(OH)D from the early follicular to mid luteal phase. Draper et al. [39] reported a significant decrease in concentration of 25(OH)D in the periovulatory and luteal phase.

Nine studies measured concentrations of 1,25(OH)<sub>2</sub>D within the follicular phase [34–38, 40–43]. Two studies found a rise in the concentrations of 1,25(OH)<sub>2</sub>D from early in the follicular phase to midcycle [35, 42]. Tjellesen et al. [42] observed a 56% increase in the concentration of 1,25(OH)<sub>2</sub>D from day 2 to day 14, and Gray et al. [35] reported a large 1,25(OH)<sub>2</sub>D increase of 128% from day 1 to 15. Conversely, Baran et al. [34] did not observe a change in concentration of 1,25(OH)<sub>2</sub>D from day 3 to day 13 of the cycle. The other six studies [36–38, 40, 41, 43] did not observe a change in 1,25(OH)<sub>2</sub>D within the follicular phase but however examined 1,25(OH)<sub>2</sub>D across the phases.

Buchanan et al. [36] found a rise of 26% from the early follicular phase to the early luteal phase. Gray et al. [35] reported a large 1,25(OH)<sub>2</sub>D increase of 128% from day 1 to 15 but in the same study a midcycle rise was *not* observed among women taking oral contraceptives. The earlier study by Thys-Jacobs et al. [40] observed a rise of 23% in 1,25(OH)<sub>2</sub>D from the early follicular phase to the late luteal phase, but failed to report statistical significance [40]; their more recent and larger study reported a modest 1,25(OH)<sub>2</sub>D increase of 13% that was statistically significant [41]. Interestingly, Muse et al. [37] measured the concentrations of 1,25(OH)<sub>2</sub>D throughout the menstrual cycle through daily blood collection but only observed a slight, non-statistically significant change across the cycle. The variation in concentrations followed no definite pattern and no midcycle rise was observed [37]. In contrast, Nielsen et al. [38] and Zittermann et al. [43] did not observe any change in concentrations of 1,25(OH)<sub>2</sub>D across the cycle.

## Discussion

After a systematic review of literature on PubMed and other databases, we identified ten longitudinal studies examining concentrations of 25(OH)D and/or 1,25(OH)<sub>2</sub>D across the normal menstrual cycle in healthy women. Five studies measured 25(OH)D, only one study reported a significant decrease in the periovulatory and luteal phase [39] and four studies found no changes in concentrations within a menstrual cycle [40–43]. Four of the nine studies measuring 1,25(OH)<sub>2</sub>D found that concentrations of 1,25(OH)<sub>2</sub>D increased across the menstrual cycle, within the follicular phase [35, 42] and/or from early to late in the cycle [36, 41]. However, five studies did not observe a change in 1,25(OH)<sub>2</sub>D concentrations across any two

cycle measurements [34, 37, 38, 40, 43], leaving an inconsistency in findings and no emerging pattern.

Our findings for 25(OH)D are similar to that of another study which examined change in concentrations of 25(OH)D during the follicular phase of the menstrual cycle among patients who were undergoing a modified natural cycle in vitro fertilization (IVF) [44]. The results suggested that concentrations of 25(OH)D, free 25(OH)D, bioavailable 25(OH)D and vitamin D binding protein (DBP) did not significantly change during the early, mid, and late follicular phase of the cycle when the estrogen levels change [44]. Two additional studies examined 25(OH)D across menstruation using a cross-sectional approach but did not test for changes in concentrations of 25(OH)D [45, 46]. Another cross-sectional study examined 25(OH)D in the early follicular phase but did not look at within phase differences [47]. Of note, one study in this review reported a decrease in 25(OH)D from the same point early in one cycle to the next, but it was presumably due to normal seasonal changes (the study was conducted from January to February, when 25(OH)D typically decreases) and was not within a cycle [43]. Only one study in this review reported a significant decrease in concentration of 25(OH)D in the periovulatory and luteal phase [39]. In the same study, concentrations of 25(OH)D were reported to be higher during menses [39]. The decrease in 25(OH)D found in this study, may be due to changes occurring in concentrations of estrogen and progesterone [48].

During the follicular phase, two out of nine studies in this review reported an increase in 1,25(OH)<sub>2</sub>D [35, 42]. We also found, in two studies out of seven that examined both menstrual phases, that concentrations of 1,25(OH)<sub>2</sub>D increased from the follicular phase to the luteal phase [36, 41]. In one of these studies, the concentration of 1,25(OH)<sub>2</sub>D was found to consistently increase from the follicular phase to luteal phase among normal women, but in the same study, a significant decrease was observed in the concentrations of 1,25(OH)<sub>2</sub>D during the early luteal phase among women with premenstrual dysphoric syndrome (PMDD) [41]. Conversely, a significantly higher concentration of 1,25(OH)<sub>2</sub>D was found during the early follicular phase among women with PMS than healthy controls [40]. A possible explanation for this change can also be related to the increased concentration of estrogen during the late follicular phase leading to a rise in 1,25(OH)<sub>2</sub>D [49]. Two of the studies which observed a change in 1,25(OH)<sub>2</sub>D, had a comparatively larger sample size which might have had an impact on the findings [36, 41]. Otherwise there were no apparent connections between studies which did and did not find changes in concentrations of 1,25(OH)<sub>2</sub>D.

Estrogens can increase the activity of 1 $\alpha$ -hydroxylase in the kidneys (the enzyme responsible for 25(OH)D to



1,25(OH)<sub>2</sub>D conversion) thereby impacting vitamin D metabolism [49]. Of note, a midcycle 1,25(OH)<sub>2</sub>D rise was not observed among women taking an oral contraceptive (albeit findings from only one study) [35], lending evidence to the hypothesis that changes in estrogens may be needed to drive 1,25(OH)<sub>2</sub>D changes. Additionally, PTH plays an important role in 1,25(OH)<sub>2</sub>D regulation and has a notable midcycle rise [50]. PTH, like estrogen, can also stimulate the production of 1,25(OH)<sub>2</sub>D by increasing the expression of renal 1 $\alpha$ -hydroxylase [50]. Overall, more research is needed to understand the determinants of 1,25(OH)<sub>2</sub>D changes, in both normal and pathological female reproductive cycles.

Reproductive hormones change across the menstrual cycle which can have an effect on changes in vitamin D metabolites across the cycle, or vice versa. The antimüllerian hormone (AMH) may have an association with 25(OH)D and thereby impact its metabolism across the menstrual cycle [51]. AMH primarily plays a role in inhibiting primordial follicle recruitment which results in a gradual decrease in follicular growth thereby delaying atresia [51, 52]. It has been suggested that vitamin D could be responsible for the regulation of AMH expression, because the AMH gene consists of a domain for the vitamin D response element in the promoter region [51]. This link could explain the apparent impact of vitamin D on ovarian function and menstrual regularity. An association has been observed between serum AMH and serum 25(OH)D among late reproductive aged women suggesting that lower concentrations of 25(OH)D can result in lower ovarian reserve [53]. 25(OH)D has also been found to be related with follicle stimulating hormone (FSH) among premenopausal women [54]. Urinary FSH is considered as a main biomarker of ovarian reserve, and a decrease in concentration of 25(OH)D has been shown to be related with higher FSH levels which can lead to reduction in primordial follicles [54]. This suggests that plasma concentrations of 25(OH)D might have an association with the ovarian reserve and thereby have an effect on women's fertility [54]. Vitamin D metabolism may also be impacted by concentrations of progesterone [48], which is higher during the luteal phase of the menstrual cycle [27]. An inverse association has been reported between 25(OH)D and progesterone suggesting that lower progesterone is related to higher 25(OH)D during the luteal phase [48]. Further research is needed to investigate mechanisms.

A final factor that we propose could be impacting concentrations of vitamin D metabolites is plasma volume, which has been shown to change across the menstrual cycle [55, 56] as well as impact concentrations of some micronutrients [29, 30, 32]. Therefore, it is currently unknown if changes in concentrations of these plasma-based biomarkers are in part due to plasma volume

changes. Overall, there are several plausible mechanisms to suggest that vitamin D metabolite concentrations might be changing across the menstrual cycle and interacting with reproductive hormones, but the exact mechanisms are yet to be elucidated.

There are many limitations to the current body of evidence; first and foremost, few studies met our longitudinal eligibility criteria and most studies had a very small sample size. Reporting was often poor with details not provided on the number of measurements or the specific means and SDs for each measurement. All studies were in high-income countries, limiting generalizability. Very few studies measured the concentration of 25(OH)D, the metabolite for which we were most interested in due to implications for understanding vitamin D status and classifying deficiency. None of the studies used liquid chromatography tandem mass spectrometry, which is considered the most valid method. Seasonal variability can influence vitamin D status which was only reported by few studies. Only one study reported details on intake of supplemental vitamin D.

## Conclusions

We found few studies that have examined vitamin D metabolites across the menstrual cycle, providing limited to insufficient data to understand potential changes or lack thereof. In the existing research, 1,25(OH)<sub>2</sub>D concentrations increased across the menstrual cycle in a few studies, but did not change in others. 25(OH)D concentrations changed across the cycle in one study but not others. Additional studies are needed to better understand 25(OH)D and 1,25(OH)<sub>2</sub>D in menstruating women. Future work should involve longitudinal studies powered to examine both metabolites (at minimum) at least two points across the menstrual cycle using the robust lab methods. Other factors that may be related to concentrations should be investigated, including the woman's age, race/ethnicity, BMI, and parity. If concentrations of 25(OH)D were found to change across the cycle, this could impact estimates of vitamin D deficiency as well as findings of associations between vitamin D status and health outcomes in women of reproductive age—both important factors in research. Vitamin D deficiency is a worldwide problem, particularly in women during reproductive years, and additional research specific to the menstrual cycle is needed to inform public health recommendations and improve research methods.

## Abbreviations

1,25(OH)<sub>2</sub>D: 1,25-dihydroxyvitamin D; 25(OH)D: 25-hydroxyvitamin D; AMH: Antimüllerian hormone; FSH: Follicle stimulating hormone; IOM: Institute of Medicine; IVF: In vitro fertilization; LH: Luteinizing hormone; LMP: Last menstrual period; PMDD: Premenstrual dysphoric syndrome; PMS: Premenstrual syndrome; PRISMA: Preferred Reporting Items for Systematic Reviews and Meta-Analysis; PTH: Parathyroid hormone; UVB: Ultraviolet B

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**Availability of data and materials**

No datasets were generated or analyzed.

**Authors' contributions**

A.D.G. conceived and designed the review with input from A.S. A.S. performed the systematic searches and wrote the manuscript. Both authors contributed to revisions of the manuscript. Both authors read and approved the final manuscript.

**Ethics approval and consent to participate**

Not applicable.

**Consent for publication**

Not applicable.

**Competing interests**

The authors declare that they have no competing interests.

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**References**

1. Looker AC, Johnson CL, Lacher DA, et al. Vitamin D Status: United States, 2001–2006. NCHS Data Brief. 2011;11–8.
2. Palacios C, Gonzalez L. Is vitamin D deficiency a major global public health problem? *J Steroid Biochem Mol Biol.* 2014;144 Pt A:138–45.
3. Mithal A, Wahl DA, Bonjour JP, et al. Global vitamin D status and determinants of hypovitaminosis D. *Osteoporos Int.* 2009;20:1807–20.
4. Ginde AA, Liu MC, CAJ C. Demographic differences and trends of vitamin D insufficiency in the US population, 1988–2004. *Arch Intern Med.* 2009;169:626–32.
5. Ross AC, Manson JE, Abrams SA, et al. The 2011 report on dietary reference intakes for calcium and vitamin D from the Institute of Medicine: what clinicians need to know. *J Clin Endocrinol Metab.* 2011;96:53–8.
6. Ginde AA, Sullivan AF, Mansbach JM, et al. Vitamin D insufficiency in pregnant and nonpregnant women of childbearing age in the United States. *Am J Obstet Gynecol.* 2010;202:436 e1–8.
7. Zhao G, Ford ES, Tsai J, et al. Factors associated with vitamin D deficiency and inadequacy among women of childbearing age in the United States. *ISRN Obstet Gynecol.* 2012;2012:691486.
8. Hossein-nezhad A, Holick MF. Vitamin D for health: a global perspective. *Mayo Clin Proc.* 2013;88:720–55.
9. Holick MF. Vitamin D deficiency. *N Engl J Med.* 2007;357:266–81.
10. Christakos S, Dhawan P, Verstuyf A, et al. Vitamin D: metabolism, molecular mechanism of action, and pleiotropic effects. *Physiol Rev.* 2016;96:365–408.
11. Holick MF, Chen TC. Vitamin D deficiency: a worldwide problem with health consequences. *Am J Clin Nutr.* 2008;87:1080S–6S.
12. Pludowski P, Holick MF, Pilz S, et al. Vitamin D effects on musculoskeletal health, immunity, autoimmunity, cardiovascular disease, cancer, fertility, pregnancy, dementia and mortality—a review of recent evidence. *Autoimmun Rev.* 2013;12:976–89.
13. Tsiaras WG, Weinstock MA. Factors influencing vitamin D status. *Acta Derm Venereol.* 2011;91:115–24.
14. Holick MF, Binkley NC, Bischoff-Ferrari HA, et al. Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab.* 2011;96:1911–30.
15. G R, Gupta A. Vitamin D deficiency in India: prevalence, causalities and interventions. *Nutrients.* 2014;6:729–75.
16. Anagnostis P, Karras S, Goulis DG. Vitamin D in human reproduction: a narrative review. *Int J Clin Pract.* 2013;67:225–35.
17. Aghajafari F, Nagulesapillai T, Ronskley PE, et al. Association between maternal serum 25-hydroxyvitamin D level and pregnancy and neonatal outcomes: systematic review and meta-analysis of observational studies. *BMJ.* 2013;346:f1169.
18. Wagner CL, Taylor SN, Dawodu A, et al. Vitamin D and its role during pregnancy in attaining optimal health of mother and fetus. *Nutrients.* 2012;4:208–30.
19. Gernand AD, Simhan HN, Baca KM, et al. Vitamin D, pre-eclampsia, and preterm birth among pregnancies at high risk for pre-eclampsia: an analysis of data from a low-dose aspirin trial. *BJOG.* 2016;124:1874–82.
20. Gernand AD, Simhan HN, Klebanoff MA, et al. Maternal serum 25-hydroxyvitamin D and measures of newborn and placental weight in a U.S. multicenter cohort study. *J Clin Endocrinol Metab.* 2013;98:398–404.
21. Gernand AD, Simhan HN, Caritis S, et al. Maternal vitamin D status and small-for-gestational-age offspring in women at high risk for preeclampsia. *Obstet Gynecol.* 2014;123:40–8.
22. Bodnar LM, Klebanoff MA, Gernand AD, et al. Maternal vitamin D status and spontaneous preterm birth by placental histology in the US collaborative perinatal project. *Am J Epidemiol.* 2014;179:168–76.
23. Luk J, Torrealday S, Neal Perry G, et al. Relevance of vitamin D in reproduction. *Hum Reprod.* 2012;27:3015–27.
24. Evans KN, Bulmer JN, Kilby MD, et al. Vitamin D and placental-decidual function. *J Soc Gynecol Investig.* 2004;11:263–71.
25. Skowronska P, Pastuszek E, Kuczynski W, et al. The role of vitamin D in reproductive dysfunction in women - a systematic review. *Ann Agric Environ Med.* 2016;23:671–6.
26. Lorenzen M, Boisen IM, Mortensen LJ, et al. Reproductive endocrinology of vitamin D. *Mol Cell Endocrinol.* 2017;453:103–12.
27. Owen JJ. Physiology of the menstrual cycle. *Am J Clin Nutr.* 1975;28:333–8.
28. Barr SI. Vegetarianism and menstrual cycle disturbances: is there an association? *Am J Clin Nutr.* 1999;70:549S–54S.
29. Michos C, Kalfakakou V, Karkabounas S, et al. Changes in copper and zinc plasma concentrations during the normal menstrual cycle in women. *Gynecol Endocrinol.* 2010;26:250–5.
30. Kim I, Yetley EA, Calvo MS. Variations in iron-status measures during the menstrual cycle. *Am J Clin Nutr.* 1993;58:705–9.
31. Kumar R, Cohen WR, Silva P, et al. Elevated 1,25-dihydroxyvitamin D plasma levels in normal human pregnancy and lactation. *J Clin Invest.* 1979;63:342–4.
32. Inoue K, Sugiyama M. Changes in body iron status, heart rate, and unspecific symptoms across menstrual cycle in marginally iron-deficient young women. *J Clin Biochem Nutr.* 1998;24:99–104.
33. Liberati A, Altman DG, Tetzlaff J, et al. The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate health care interventions: explanation and elaboration. *PLoS Med.* 2009;6:e1000100.
34. Baran DT, Whyte MP, Haussler MR, et al. Effect of the menstrual cycle on calcium-regulating hormones in the normal young woman. *J Clin Endocrinol Metab.* 1980;50:377–9.
35. Gray TK, McAdoo T, Hatley L, et al. Fluctuation of serum concentration of 1,25-dihydroxyvitamin D<sub>3</sub> during the menstrual cycle. *Am J Obstet Gynecol.* 1982;144:880–4.
36. Buchanan JR, Santen RJ, Cavaliere A, et al. Interaction between parathyroid hormone and endogenous estrogen in normal women. *Metabolism.* 1986; 35:489–94.
37. Muse KN, Manolagas SC, Deftos LJ, et al. Calcium-regulating hormones across the menstrual cycle. *J Clin Endocrinol Metab.* 1986;62:1313–6.
38. Nielsen HK, Brixen K, Bouillon R, et al. Changes in biochemical markers of osteoblastic activity during the menstrual cycle. *J Clin Endocrinol Metab.* 1990;70:1431–7.
39. Draper CF, Duisters K, Weger B, et al. Menstrual cycle rhythmicity: metabolic patterns in healthy women. *Sci Rep.* 2018;8:14568.
40. Thys-Jacobs S, Alvir MJ. Calcium-regulating hormones across the menstrual cycle: evidence of a secondary hyperparathyroidism in women with PMS. *J Clin Endocrinol Metab.* 1995;80:2227–32.
41. Thys-Jacobs S, McMahon D, Bilezikian JP. Cyclical changes in calcium metabolism across the menstrual cycle in women with premenstrual dysphoric disorder. *J Clin Endocrinol Metab.* 2007;92:2952–9.
42. Tjellesen L, Christiansen C, Hummer L, et al. Unchanged biochemical indices of bone turnover despite fluctuations in 1,25-dihydroxyvitamin D during the menstrual cycle. *Acta Endocrinol (Copenh).* 1983;102:476–80.
43. Zittermann A, Schwarzl I, Scheld K, et al. Physiologic fluctuations of serum estradiol levels influence biochemical markers of bone resorption in young women. *J Clin Endocrinol Metab.* 2000;85:95–101.

44. Franasiak JM, Wang X, Molinaro TA, et al. Free vitamin D does not vary through the follicular phase of the menstrual cycle. *Endocrine*. 2016;53:322–6.
45. Jukic AM, Steiner AZ, Baird DD. Lower plasma 25-hydroxyvitamin D is associated with irregular menstrual cycles in a cross-sectional study. *Reprod Biol Endocrinol*. 2015;13:20.
46. Jukic AM, Upson K, Harmon QE, et al. Increasing serum 25-hydroxyvitamin D is associated with reduced odds of long menstrual cycles in a cross-sectional study of African American women. *Fertil Steril*. 2016;106:172–9 e2.
47. Lagowska K. The relationship between vitamin D status and the menstrual cycle in young women: a preliminary study. *Nutrients*. 2018;10.
48. Knight JA, Wong J, Blackmore KM, et al. Vitamin D association with estradiol and progesterone in young women. *Cancer Causes Control*. 2010;21:479–83.
49. Buchanan JR, Santen R, Cauffman S, et al. The effect of endogenous estrogen fluctuation on metabolism of 25-hydroxyvitamin D. *Calcif Tissue Int*. 1986;39:139–44.
50. Fraser DR, Kodicek E. Regulation of 25-hydroxycholecalciferol-1-hydroxylase activity in kidney by parathyroid hormone. *Nat New Biol*. 1973;241:163–6.
51. Irani M, Merhi Z. Role of vitamin D in ovarian physiology and its implication in reproduction: a systematic review. *Fertil Steril*. 2014;102:460–8 e3.
52. Peluso C, Fonseca FL, Rodart IF, et al. AMH: an ovarian reserve biomarker in assisted reproduction. *Clin Chim Acta*. 2014;437:175–82.
53. Merhi ZO, Seifer DB, Weedon J, et al. Circulating vitamin D correlates with serum antimullerian hormone levels in late-reproductive-aged women: Women's interagency HIV study. *Fertil Steril*. 2012;98:228–34.
54. Jukic AM, Steiner AZ, Baird DD. Association between serum 25-hydroxyvitamin D and ovarian reserve in premenopausal women. *Menopause*. 2015;22:312–6.
55. Cullinane EM, Yurgalevitch SM, Saritelli AL, et al. Variations in plasma volume affect total and low-density lipoprotein cholesterol concentrations during the menstrual cycle. *Metabolism*. 1995;44:965–71.
56. Bernstein IM, Ziegler W, Badger GJ. Plasma volume expansion in early pregnancy. *Obstet Gynecol*. 2001;97:669–72.

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